



Research Article

Algae 2022, 37(2): 123-133

<https://doi.org/10.4490/algae.2022.37.6.4>

Open Access



Phylogeographic patterns in cryptic *Bostrychia tenella* species (Rhodomelaceae, Rhodophyta) across the Thai-Malay Peninsula

Jakaphan Bulan¹, Sinchai Maneekat¹, Giuseppe C. Zuccarello² and Narongrit Muangmai^{1,*}

¹Department of Fishery Biology, Faculty of Fisheries, Kasetsart University, Bangkok 10900, Thailand

²School of Biological Science, Victoria University of Wellington, P.O. Box 600, Wellington 6140, New Zealand

Genetic diversity and distribution patterns of marine macroalgae are increasingly being documented in Southeast Asia. These studies show that there can be significant levels of genetic diversity and isolation between populations on either side of the Thai-Malay Peninsula. *Bostrychia tenella* is a common filamentous red seaweed in the region and the entity is represented by at least two cryptic species. Despite being highly diverse and widespread, genetic variation and population structure of this species complex remains understudied, especially around the Thai-Malay Peninsula. We analyzed genetic diversity and inferred the phylogeographic pattern of specimens identified as *B. tenella* using the plastid RuBisCo spacer from samples from the Andaman Sea and the Gulf of Thailand. Our genetic analysis confirmed the occurrence of the two cryptic *B. tenella* species (B and C) along both coasts. Cryptic species B was more common in the area and displayed higher genetic diversity than species C. Historical demographic analyses indicated a stable population for species B, but more recent population expansion for species C. Our analyses also revealed that both cryptic species from the Andaman Sea possessed higher genetic diversity than those of the Gulf of Thailand. We also detected moderate to high levels of gene flow and weak phylogeographic structure of cryptic species B between the two coasts. In contrast, phylogeographic analysis showed genetic differences between populations of both cryptic species within the Andaman Sea. Overall, these results suggest that cryptic *B. tenella* species around Thai-Malay Peninsula may have undergone different demography histories, and their patterns of genetic diversity and phylogeography were likely caused by geological history and regional sea surface current circulation in the area.


Keywords: connectivity; cryptic diversity; dispersal; macroalgae; Sundaland

INTRODUCTION

Genetic studies are increasingly exploring the patterns of diversity, evolutionary history, and phylogeography for many marine species (Bowen et al. 2016, Faria et al. 2021). Phylogeographic analyses are often used to trace the process of marine species dispersal and population connectivity (e.g., Zuccarello and Martin 2016, Vieira et al. 2021), but can also detect cryptic species diversity and biogeographic breaks in the ocean (Fraser et al. 2013,

Muangmai et al. 2015a). Such analyses are crucial to provide a useful framework for marine natural bioresource management and invasive species risk assessment (Teske et al. 2011, Bowen et al. 2016).

In the Southeast Asia region, genetic diversity and phylogeographic patterns of macroalgae are being increasingly documented. Many previous studies indicated different levels of genetic diversity among macroalgal species.

 This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Received February 1, 2022, Accepted June 4, 2022

*Corresponding Author

E-mail: ffisnrm@ku.ac.th

Tel: +66-2-942-8701, Fax: +66-2-942-8702

For example, the brown alga *Sargassum polycystum* C. Agardh showed low genetic diversity, but higher diversity on the Andaman coast of Thailand (Kantachumpoo et al. 2014), whereas, in the Philippines, high genetic diversity was detected in the red algae, *Phycocalidia acanthophora* (E. C. Oliveira & Coll) Santiañez (Dumilag and Aguinaldo 2017, as *Pyropia acanthophora*) and *Gracilaria salicornia* (C. Agardh) E. Y. Dawson (Ferrer et al. 2019). Phylogeographic studies also indicated partial isolation between the Indian and Pacific Ocean populations of *S. polycystum* (Kantachumpoo et al. 2014). This separation appears to be maintained by a geographic barrier (the Thai-Malay Peninsula) and the current oceanographic discontinuity between the Indian and Pacific oceans (Kantachumpoo et al. 2014, Wichachucherd et al. 2014).

Cryptic species are common in red algae (e.g., Zuccarello et al. 2002, Muangmai et al. 2014, Díaz-Tapia et al. 2018) and are known to differ in many species-specific properties, although being morphologically identical. Their demographic histories can differ (Muangmai et al. 2015a, 2022) and they can differ physiologically (Muangmai et al. 2015b) and chemically (Payo et al. 2011, Bracegirdle et al. 2019). In addition, they can differ in their intertidal position (Muangmai et al. 2016). In the Indo-Pacific, genetic studies revealed cryptic diversity in many red seaweeds (e.g., Payo et al. 2013, Gabriel et al. 2016, Saengkaew et al. 2016). Recognizing this cryptic algal diversity highlights a substantial undiscovered biodiversity, with implication for a need to continue a thorough investigation of red algae using data based on extensive markers and sampling.

Bostrychia tenella (J. V. Lamouroux) J. Agardh is a marine filamentous red alga, which is distributed in tropical and subtropical regions (King and Puttock 1989, Zuccarello et al. 2015). This species naturally grows either on mangroves or in the high intertidal on rocks. Zuccarello et al. (2015) were able to resurrect the species *B. binderi* Harvey from *B. tenella* using DNA-based species delimitation, but also showed cryptic species diversity within *B. tenella*, consisting of two species B and C with overlapping distributions. In Thailand, *B. tenella* is relatively common and widespread across the coastal areas based on morphological record (Lewmanomont et al. 1995). Recent genetic analysis indicated the occurrence of *B. tenella* species B and C on both eastern (the Gulf of Thailand - Pacific Ocean) and western coasts (Andaman Sea - Indian Ocean) of Thailand (Saengkaew et al. 2016). Additionally, coexistence of two cryptic species was reported from both coastal regions of the country (Saengkaew et al. 2016). However, due to the limited sampling this study

mainly focused on the phylogenetic diversity of *B. tenella*, and therefore genetic diversity and distribution pattern of this cryptic species across the Thai-Malay Peninsula have not been fully examined.

In this present study, we used the chloroplast-encoded RuBisCo spacer of *B. tenella* across the Thai-Malay Peninsula to assess its genetic diversity and phylogeographic pattern. We also wanted to see if there were genetic differences of these cryptic species between the Andaman coasts and the Gulf of Thailand, and then determined what factors could explain these distribution patterns.

MATERIALS AND METHODS

Algal sampling

We surveyed *B. tenella* populations around rocky intertidal areas of 12 sites along the Thai-Malay Peninsula: the Gulf of Thailand (T1-T6) and the Andaman Sea (A1-A6) (Table 1, Fig. 1), between 2017 and 2019. At each site, algal samples were randomly collected, at least 10 samples per population, and preserved in silica gel for both morphological and molecular analyses. For morphological observation, preserved samples were prepared by soaking in water for 5 min and subsequently simultaneously stained and preserved in 1% aniline blue acidified with 1% HCl and mounted in 50% glucose syrup (Karo Corn Syrup) on microscope slides. Species identification was based on previous publications (King and Puttock 1989, Zuccarello et al. 2015).

DNA extraction, polymerase chain reaction, and sequencing

Genomic DNA isolation was performed using apical portions of dried algal specimens, with a modified Chelex extraction method (Zuccarello et al. 1999) or Norgen Biotek's Plant/Fungi DNA Isolation Kit (Norgen Biotek Corp., Thorold, Canada). The plastid-encoded RuBisCo spacer was selected for polymerase chain reaction (PCR) amplification. This fragment is a powerful region for exploring inter- and intra-species diversity in *Bostrychia* (Zuccarello and West 2003, Zuccarello et al. 2015). PCR was carried out using PCR Master mix Solution (i-Taq, iNtRON Biotechnology DR, Seongnam, Korea), in a total volume of 20 µL, consisting of 10 µL of i-Taq, 10 pmol of each primer and 2 µL of genomic DNA (~10–20 ng). PCR amplification profile and procedure followed Zuccarello and West (2003). All amplified products were cleaned

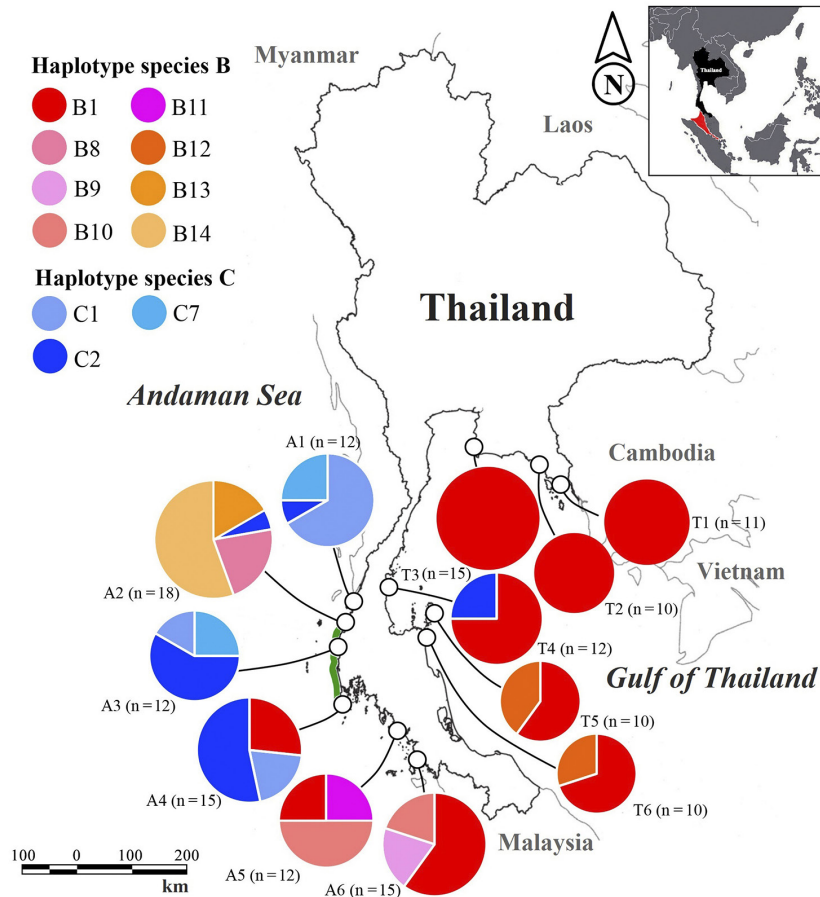


Fig. 1. Distribution of *Bostrychia tenella* species B and C based on RuBisCo spacer haplotypes across the Thai-Malay Peninsula. Pie charts on the map indicate the relative proportions of haplotypes at each population (Table 1), with the total number of samples (n) per population given next to each pie chart. Green line indicates possible phylogeographic break zone within the Andaman Sea. (Inset) Map of Southeast Asia showing Thailand shaded in black and Malacca Strait shaded in red.

using ExoSAP-IT (USB, Cleveland, OH, USA) and then sequenced commercially in both directions using PCR primers (U2Bio Inc., Seoul, Korea).

Alignment, phylogenetic reconstructions, historical demography, and population structure analysis

Sequences were edited, assembled, and aligned using the Geneious Prime software package (Biomatters, available from <http://www.geneious.com/>). Alignments of the RuBisCo spacer sequences were performed using the MAFFT algorithm (Kato et al. 2002) and were further manually refined. Additional 12 sequences of *B. tenella* were obtained from GenBank and included in the alignment (Supplementary Table S1).

Phylogenetic trees were reconstructed using maximum-likelihood (ML) and Bayesian inference (BI) meth-

ods using IQ-TREE (Minh et al. 2020) and MrBayes v3.2 (Ronquist et al. 2012), respectively. ML analyses were carried out with the HKY + G model as selected by ModelFinder (Kalyaanamoorthy et al. 2017) and 1,000 bootstrap replicates. The molecular evolution models for BI were selected using Kakusan 4 (Tanabe 2011). BI analyses were performed using a GTR + I + R model, with two parallel runs of four Markov chains for a million generations. We sampled one tree every 1,000 generations and then removed 2,500 trees (burn-in) before determining a consensus tree and posterior probabilities. Both ML and BI trees were edited with the program FigTree v1.4.4 (Rambaut 2016).

Number of haplotypes (H), haplotype (Hd), and nucleotide (π) diversity were calculated for each population using DnaSP v6 (Rozas et al. 2017). Haplotype networks were produced using median joining generated in PopART v1.7 (University of Otago, available from [125](http://</p>
</div>
<div data-bbox=)

Table 1. Collection localities and genetic diversity parameters of cryptic *Bostrychia tenella* species in Thailand

Population code	Locality (collection date)	Coordinates	No. of individuals examined (n)	Haplotype present	No. of haplotypes [H (n)]	Haplotype diversity (Hd ± SD)	Nucleotide diversity (π ± SD)
Andaman Sea							
A1	Chandamri Beach, Ranong (Mar 2017)	9°57'57.2" N, 98°35'34.5" E	12	C = 3	C1 (8), C2 (1), C7 (3)	C = 0.53 ± 0.13	C = 0.00236 ± 0.0001
A2	Prapas Beach, Ranong (Mar 2017)	9°21'45.8" N, 98°23'41.9" E	18	B = 3 C = 1	B8 (3), B13 (4), B14 (10) C2 (1)	B = 0.603 ± 0.09 C = 0.00	B = 0.00369 ± 0.0001 C = 0.00
A3	Ao Khoei, Phang Nga (Mar 2017)	9°17'19.8" N, 98°22'21.7" E	12	C = 3	C1 (3), C2 (7), C7 (2)	C = 0.621 ± 0.11	C = 0.00293 ± 0.0006
A4	Saphan Hin Beach, Phuket (Apr 2018)	7°52'06.8" N, 98°23'58.3" E	15	B = 1 C = 2	B1 (4) C1 (3), C2 (8)	B = 0.00 C = 0.436 ± 0.13	B = 0.00 C = 0.00153 ± 0.0004
A5	Ratchamongkol Beach, Trang (Mar 2018)	7°31'15.8" N, 99°18'19.8" E	12	B = 3	B1 (3), B10 (6), B11 (3)	B = 0.682 ± 0.09	B = 0.00286 ± 0.0005
A6	Pak Bara, Satun (Mar 2018)	6°51'41.4" N, 99°43'11.1" E	15	B = 3	B1 (9), B9 (3), B10 (3)	B = 0.600 ± 0.10	B = 0.00360 ± 0.00001
Total			84	B = 7 C = 3	B1 (16), B8 (3), B9 (3), B10 (9), B11 (3), B13 (4), B14 (10) C1 (14), C2 (17), C7 (5)	B = 0.829 ± 0.02 C = 0.629 ± 0.04	B = 0.01369 ± 0.0003 C = 0.00272 ± 0.0001
Gulf of Thailand							
T1	Laem Sok, Trat (May 2019)	12°02'47.3" N, 102°35'10.3" E	11	B = 1	B1 (11)	B = 0.00	B = 0.00
T2	Kung Krabaen Bay, Chanthaburi (May 2019)	12°31'24.5" N, 101°56'56.4" E	10	B = 1	B1 (10)	B = 0.00	B = 0.00
T3	Samaesan, Chonburi (Mar 2018)	12°36'09.9" N, 100°57'08.9" E	15	B = 1	B1 (15)	B = 0.00	B = 0.00
T4	Thungwualaen Beach, Chumphon (Apr 2019)	10°33'15.4" N, 99°16'38.1" E	12	B = 1 C = 1	B1 (9) C2 (3)	B = 0.00 C = 0.00	B = 0.00 C = 0.00
T5	Samui Island, Surat Thani (Apr 2019)	9°25'32.4" N, 99°56'05.0" E	10	B = 2	B1 (7), B12 (3)	B = 0.533 ± 0.09	B = 0.00374 ± 0.00001
T6	Khanom, Nakhon Si Thammarat (Apr 2019)	9°08'15.1" N, 99°52'38.7" E	10	B = 2	B1 (6), B12 (4)	B = 0.467 ± 0.13	B = 0.00327 ± 0.00001
Total			68	B = 2 C = 1	B1 (58), B12 (7) C2 (3)	B = 0.195 ± 0.06 C = 0.00	B = 0.00137 ± 0.00042 C = 0.00
Overall			152	B = 8 C = 3	B1 (16), B8 (3), B9 (3), B10 (9), B11 (3), B12 (7), B13 (4), B14 (10) C1 (14), C2 (20), C7 (5)	B = 0.670 ± 0.05 C = 0.629 ± 0.04	B = 0.00807 ± 0.00021 C = 0.00127 ± 0.00016

SD, standard deviation.

popart.otago.ac.nz.). Additionally, the historical demography for each species (species B vs. species C) and population (Andaman Sea vs. the Gulf of Thailand) levels was estimated by the analyses of mismatch distribution and neutrality tests, Tajima's D (Tajima 1989) and Fu's F_s , as implemented in DnaSP. The analysis of mismatch distribution uncovers the demographic history of populations, with population expansions producing a unimodal distribution, whereas stable population show a multimodal distribution (Rogers and Harpending 1992).

Population differentiation was analyzed separately for each cryptic species for populations with at least 10 samples. The analysis was examined using two different approaches. First, pairwise population differentiation (F_{ST}) values were estimated with a significance level of 0.05 determined by 10,000 permutations. Secondly, Analysis of Molecular Variance (AMOVA) was performed with significance determined by 1,023 permutations. All analyses were conducted in Arlequin v 3.5.1.3 (Excoffier and Lischer 2010).

RESULTS

Genetic diversity and distribution

The RuBisCo spacer with partial 3'-*rbcl* and 5'-*rbcs* gene sequences of 275–286 bp were successfully generated from 152 samples of 12 populations of *B. tenella* from both coasts of Thailand. Genetic distance among these sequences ranged from 0.0 to 6.6%. ML and BI analyses yielded almost complete topological congruence, and the ML tree is presented in Supplementary Fig. S1. Phylogenetic analyses demonstrated the occurrence of two different cryptic species of *B. tenella*: B and C, based on Zuccarello et al. (2015), in Thailand. Among 152 samples, 113 samples were species B, while 39 samples belonged to species C. Intraspecific genetic variation varied from 0.0–3.4% for species B and ranged from 0.0–1.3% for species C.

Haplotype and genetic diversity indices of the two cryptic species are presented in Table 1. Within species, both cryptic *B. tenella* species revealed a moderate genetic diversity ($Hd = 0.670$ for species B and $Hd = 0.629$ for species C), but higher nucleotide diversity for species B ($\pi = 0.00807$) than for species C ($\pi = 0.00272$) (Table 1). Within populations haplotype diversity (Hd) ranged from 0.00 to 0.68 for species B and from 0.00 to 0.62 for species C (Table 1). Nucleotide diversity (π) was relatively low, varying from 0.0000 to 0.0037 for species B, and from

0.0000 to 0.0023 for species C (Table 1).

Both cryptic *B. tenella* species were found on both coastal areas of Thailand. Cryptic species B was widely distributed in Thailand, recorded in 9 out of 12 sampled populations, while species C was detected in 7 populations (Fig. 1). Additionally, both cryptic species were found to coexist at three of the 12 sites (A2 and A4 in the Andaman Sea and T4 from the Gulf of Thailand) (Fig. 1).

Haplotype diversity and networks

Median-joining haplotype networks of cryptic species B and C constructed using RuBisCo spacer sequence (including haplotypes from Zuccarello et al. 2015 and Saengkaew et al. 2016) are presented in Fig. 2. Cryptic species B consisted of eight haplotypes in Thailand (B1, B8–B14) of which the B9–B14 are reported here for the first time. The haplotype network of species B was star-like with a central common haplotype B1 (80% of samples), found in nine of 12 populations sampled in the present study (Table 1, Fig. 1) and in addition at three sampled sites (Phuket, Chumphon and Nakhon Si Thammarat) of Saengkaew et al. (2016) and in Sabah, Malaysia (Zuccarello et al. 2015). Five novel B-haplotypes were only found in Andaman populations (A2, A5, and A6), while haplotype B12 was restricted to the Gulf of Thailand (populations T5 and T6) (Table 1, Fig. 1).

Cryptic species C is represented by three haplotypes in Thailand (C1, C2, and C7) (Fig. 2). Haplotype C2 was relatively common, accounting for 51% of all samples, and it occurred in all five populations where this species was found. C2 is the only haplotype of cryptic species C which was shared on both coasts of Thailand, while the other haplotypes were confined to the Andaman Sea (Table 1, Fig. 1). Haplotype C1 was shared in nearly all population of this species in the Andaman Sea, while haplotype C7 was detected from populations A1 and A3 (Table 1, Fig. 1).

Demographic history and population differentiation

The mismatch distribution showed a multimodal pairwise distance distribution of species B, but unimodal distribution of species C, indicating that species C has undergone a recent expansion (Supplementary Fig. S2). These demographic patterns were further supported by significant positive values of Tajima's D test for species B (0.08932, $p < 0.05$), suggesting a stable population, but significant negative value for species C (-1.11856, $p < 0.05$), indicating a population expansion. However, the

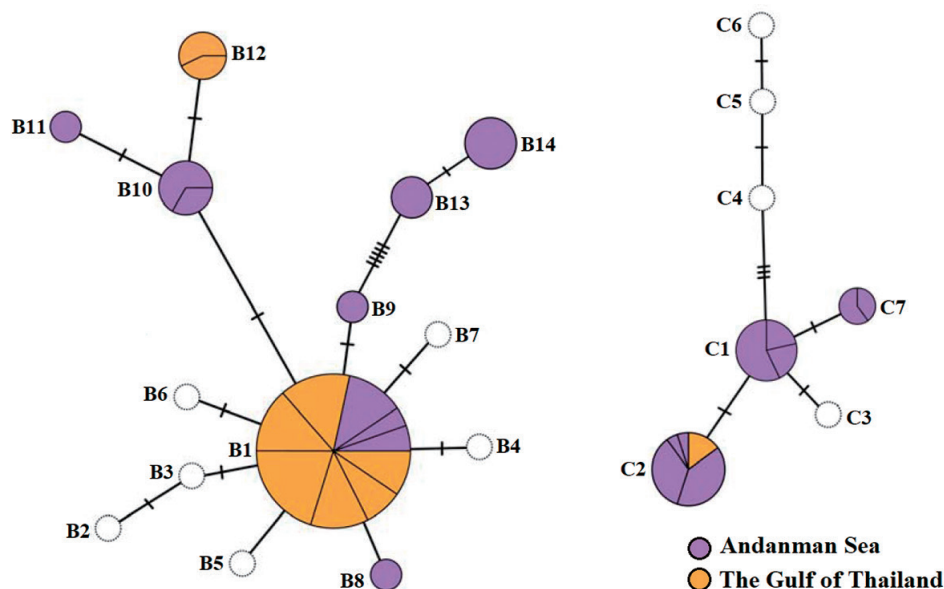


Fig. 2. Haplotype networks for cryptic *Bostrychia tenella* species B and C from RuBisCo spacer sequences. Colors represent the populations in the Andaman Sea (purple) and in the Gulf of Thailand (orange). Pie chart size represents the frequency of each haplotype. Solid circles correspond to haplotypes found in this study, whereas dashed and white colored circles correspond to haplotypes from Zuccarello et al. (2015). Small lines represent inferred missing haplotypes.

estimates of Fu's F_s were not significant (1.12591, $p > 0.1$ for species B, and 1.02017, $p > 0.1$ for species C).

Analyses within biogeographic regions were only done for *B. tenella* species B. Species B from the Andaman Sea displayed a multimodal distribution (Supplementary Fig. S2) and significant positive value of Fu's F_s test (1.78007, $p < 0.05$), whereas species B from the Gulf of Thailand showed a unimodal distribution (Supplementary Fig. S2) plus negative values of Fu's F_s test (-0.54417, $p > 0.1$), implying that the Gulf populations has more likely undergone recent expansion than the Andaman Sea populations. Values of Tajima's D test were not significant (1.74664, $p > 0.1$ for Andaman Sea population and 1.34228, $p > 0.1$ for the Gulf population).

Genetic differentiation (F_{ST} values) was moderate to low between populations of *B. tenella* species B from the Andaman Sea (A2, A5, and A6) and the Gulf of Thailand (T1-T6), with pairwise F_{ST} values ranging between 0.14 and 0.69 ($p < 0.05$) (Supplementary Table S2). Significant moderate to low genetic differentiation was detected within Andaman Sea populations, with F_{ST} value in the range of 0.67 of 0.28 (A2, A5, and A6). While low genetic differentiation was discovered from the Gulf of Thailand population, with F_{ST} value in the range of 0.08 to 0.40 (Supplementary Table S2). No genetic differentiation ($F_{ST} = 0.00$) was found among the upper Gulf populations (T1-T4), while a slight genetic differentiation ($F_{ST} = 0.08$) was

observed between the lower Gulf populations (T5 and T6) (Supplementary Table S2).

Pairwise F_{ST} values indicated that Andaman Sea populations of cryptic species C differed significantly. Population A1 was genetically distinct from the other two populations, with F_{ST} values ranging between 0.40 to 0.51. Low genetic differentiation was observed between populations A3 and A4 ($F_{ST} = 0.03$) (Supplementary Table S3).

The AMOVA was calculated to determine genetic differentiation of cryptic species B between the Andaman Sea (A2, A5, and A6) and the Gulf of Thailand (T1-T6) populations. AMOVA results indicated that the proportion of variation attributed to among populations within regions was 47.70% and within-population differences for 32.43%, whereas 19.87% occurred among regions (Supplementary Table S4). The F statistics revealed the significant and high genetic differences within populations ($F_{ST} = 0.67$), but low among regions ($F_{CT} = 0.19$) (Supplementary Table S4).

DISCUSSION

With wide-range sampling along the coasts of Thailand and chloroplast DNA analysis, we reconfirmed the occurrence of two distinct lineages, B and C, of *B. tenella* on both coastal regions of the country. In addition, we de-

ected differences between these two cryptic species. *Bostrychia tenella* species B showed higher abundance and genetic diversity than species C. Different levels of genetic diversity between cryptic species was also observed in other red seaweeds, e.g., *Bostrychia intricata* (Bory Montagne (Muangmai et al. 2015a, 2022), *Sarcopeltis skottsbergii* (Setchell & N. L. Gardner) Hommersand, Hughey, Leister & P. W. Gabrielson (Billard et al. 2015, as *Gigartina skottsbergii*), and *Asparagopsis taxiformis* (Delile) Trevisan (Zanolla et al. 2018). These previous studies suggested that these different levels of genetic variation among cryptic species was likely due to different evolutionary and demographic histories. Our additional analyses of mismatch distribution and Tajima's D test further supported the different patterns of historic population demography in these two cryptic species of *B. tenella*. We, therefore, postulate that cryptic *B. tenella* species B and C in this area have different patterns of historical population demography. However, the evolutionary history of cryptic *B. tenella* species needs further investigation with larger sample sizes and more variable markers (e.g., microsatellites).

We also observed different levels of genetic diversity in cryptic *B. tenella* species between the Andaman Sea and the Gulf of Thailand. Our data showed that genetic diversity of Andaman Sea populations of *B. tenella* was higher than that of the populations in the Gulf of Thailand. Additionally, we identified several novel haplotypes of cryptic species B from the Andaman Sea, whereas only one new haplotype was found in the Gulf of Thailand. This is consistent with previous findings that the Andaman Sea harbored greater diversity, both genetic and at the species levels than the Gulf of Thailand for seaweed (e.g., Kantachumpoo et al. 2014, Wichachucherd et al. 2014, Pongparadon et al. 2015) and other marine species, for example littoral earthworms (Seesamut et al. 2019), dugongs (Poommouang et al. 2021) and clams (Suppapan et al. 2021). Differences in genetic diversity across the Thai-Malay Peninsula could be driven by historical environmental changes and different geographical features. The lowering of sea levels (~120 m below present-day sea level) during the Last Glacial Period (ca. 17,000 years ago) resulted in the formation of landmass in Southeast Asia known as Sundaland (Sathiamurthy and Voris 2006). The emergent Sundaland would have eliminated the habitat availability of marine taxa through the Gulf of Thailand and constrained populations to the east (South China Sea) and west (the Andaman Sea) coasts of the landmass (Cannon et al. 2009, Ludt and Rocha 2015). By the end of the last glaciation (ca. 11,000 years ago), recoloniza-

tion of the Gulf of Thailand would have led to a potential founder event for these populations (Palmer 2004, Guo et al. 2020). Accordingly, this would result in higher genetic diversity in Andaman Sea populations, and lower diversity in the Gulf population. Such a hypothesis has been applied to mangrove species in Southeast Asia (see Wee et al. 2014, Guo et al. 2020), and therefore could also apply to *B. tenella* and other seaweed around the Thai-Malay Peninsula.

Although previous studies have revealed strong phylogeographic patterns between the Gulf of Thailand and Andaman Sea in some marine species (e.g., Nguyen et al. 2014, Wichachucherd et al. 2014, Seesamut et al. 2019, Panithanarak 2020), we did not find a clear phylogeographic signal in the *B. tenella* species complex across the Thai-Malay Peninsula. Our results showed that pairwise F_{ST} value between the Andaman Sea and the Gulf populations of cryptic species B demonstrated the presences of low to moderate levels of genetic differentiation, with only 19.87% of detected genetic differences found between the Andaman Sea and the Gulf of Thailand (AMOVA results), indicating moderate to high connectivity in cryptic species B. Our findings suggest the occurrence of gene flow around the Thai-Malay Peninsula. Furthermore, our demographic history analyses indicated that the Gulf populations of cryptic species B historically experienced a more recent expansion than Andaman Sea populations. Accordingly, we assume that the connectivity of *B. tenella* species B population across Thai-Malay Peninsula could possibly be associated with the historical subsidence of Sundaland, leading to a recent population expansion of this alga during the inundation of the Gulf of Thailand, and regional circulation patterns linking these two coastal areas (Haditjar et al. 2020).

In the Andaman Sea, we detected some genetic differences between *B. tenella* species. Cryptic species B population from Ranong province (A2) differed significantly from Trang and Satun provinces (A5 and A6). Additionally, analysis of haplotype distribution indicated that haplotype B1 found in population A4 was also found in A5 and A6, but not in A2, implying the genetic break occurring north to Phuket provinces (A4) (Fig. 1). Similarly, pairwise F_{ST} showed moderate genetic differentiation of cryptic species C populations between Ranong province (A1) and Phang Nga and Phuket provinces (A3 and A4). These results indicate that there is low gene flow between these populations of both cryptic species. Potential factors contributing to the genetic differentiation could be attributed to the ocean circulation patterns from North Indian Ocean and the Malacca Strait (Wyrski 1961, Chat-

terjee et al. 2017). The North Indian Ocean current always flows southwesterly toward the Andaman Sea, while the Malacca Strait current directs north-westward toward the Andaman Sea. These two currents flowed offshore and partly mix during both the North–East (December through February) and the South–West (June through September) monsoon season (Rizal et al. 2012). This current pattern may act as a marine barrier to gene flow, which promotes isolation between Andaman Sea populations. Such potential barrier was speculated to contribute to restricted distribution of some *Halimeda* species (Pongparadon et al. 2015) and genetic divergence of sandfish (Ninwichian and Klinbunga 2020) and mangrove (Guo et al. 2020).

Our results also showed the coexistence of cryptic *B. tenella* species B and C in populations of the Andaman Sea and the Gulf of Thailand. Coexistence of different cryptic species is likely to be common in the genus *Bostrychia* (Zuccarello and West 2003, 2011, Muangmai et al. 2014, 2022). How different cryptic *Bostrychia* species can coexist could possibly be explained by their eco-physiological differences, including difference in tidal height distribution, wave exposure and salinity tolerance (Muangmai et al. 2015b, 2016). In order to obtain detailed insight into the forces maintaining the coexistence in cryptic *B. tenella* species, their ecological interactions and physiological aspects need further investigation.

In conclusion, our data clearly indicated the contrasting patterns of genetic diversity between two cryptic *B. tenella* species B and C in Thai waters, suggesting different demographic history between these two cryptic species. However, both cryptic species from Andaman Sea possessed higher level of genetic diversity than the Gulf of Thailand populations. These findings have implications for not only evaluating the potential genetic resources of these cryptic red algae, but even contributing to accurate marine biodiversity assessment for strategic conservation planning of coastal areas. We also detected weak phylogeographic structure of *B. tenella* species B between the two coasts, but a relatively strong phylogeographic patterning of *B. tenella* species C along the Andaman Sea coast. These patterns could have been shaped by a combination of historical geological events (submergence of Sundaland) and contemporary regional circulation. However, in order to better understand the phylogeographic structure and demographic history of cryptic *B. tenella* species distributed around the Thai-Malay Peninsula, additional sampling sites from the Malacca Strait and the South China Sea, plus applying more variable genetic markers are necessary.

ACKNOWLEDGEMENTS

This study is funded by Kasetsart University through the Graduate School Fellowship Program (JB and NM), and Faculty of Fisheries, Kasetsart University (NM). We also appreciate Jantana Praiboon and Chatcharee Kaewsuralikhit for assistance with algal sample collection.

CONFLICTS OF INTEREST

The authors declare that they have no potential conflicts of interest.

SUPPLEMENTARY MATERIALS

Supplementary Table S1. RuBisCo spacer sequence of *Bostrychia* samples used in this study (<https://www.e-algae.org>).

Supplementary Table S2. Pairwise F_{ST} values for the 12 populations of *Bostrychia tenella* species B along the coast of Thailand (<https://www.e-algae.org>).

Supplementary Table S3. Pairwise F_{ST} values for the 12 populations of *Bostrychia tenella* species C along the Andaman coasts of Thailand (<https://www.e-algae.org>).

Supplementary Table S4. Analysis of molecular variance (AMOVA) results of *Bostrychia tenella* species B (<https://www.e-algae.org>).

Supplementary Fig. S1. The maximum-likelihood (ML) phylogeny ($\ln L = -2,325.1355$) of *Bostrychia tenella* inferred from plastid RuBisCo spacer sequences showing the two cryptic species (B and C, from Zuccarello et al. 2015) (<https://www.e-algae.org>).

Supplementary Fig. S2. Mismatch distributions of the cryptic *B. tenella* species B (A) and species C (B). Mismatch distributions of species B from the Andaman Sea (C) and from the Gulf of Thailand (D) (<https://www.e-algae.org>).

REFERENCES

- Billard, E., Reyes, J., Mansilla, A., Faugeron, S. & Guillemin, M. -L. 2015. Deep genetic divergence between austral populations of the red alga *Gigartina skottsbergii* reveals a cryptic species endemic to the Antarctic continent. *Polar Biol.* 38:2021–2034.
- Bowen, B. W., Gaither, M. R., DiBattista, J. D., Iacchei, M., Andrews, K. R., Grant, W. S., Toonen, R. J. & Briggs, J. C.

2016. Comparative phylogeography of the ocean planet. *Proc. Natl. Acad. Sci. U. S. A.* 113:7962–7969.
- Bracegirdle, J., Sohail, Z., Fairhurst, M. J., Gerth, M. L., Zuccarello, G. C., Hashmi, M. A. & Keyzers, R. A. 2019. Costatone C: a new halogenated monoterpene from the New Zealand red alga *Plocamium angustum*. *Mar. Drugs* 17:418.
- Cannon, C. H., Morley, R. J. & Bush, A. B. G. 2009. The current refugial rainforests of Sundaland are unrepresentative of their biogeographic past and highly vulnerable to disturbance. *Proc. Natl. Acad. Sci. U. S. A.* 106:11188–11193.
- Chatterjee, A., Shankar, D., McCreary, J. P., Vinayachandran, P. N. & Mukherjee, A. 2017. Dynamics of Andaman Sea circulation and its role in connecting the equatorial Indian Ocean to the Bay of Bengal. *J. Geophys. Res. Oceans* 122:3200–3218.
- Díaz-Tapia, P., Maggs, C. A., Macaya, E. C. & Verbruggen, H. 2018. Widely distributed red algae often represent hidden introductions, complexes of cryptic species or species with strong phylogeographic structure. *J. Phycol.* 54:829–839.
- Dumilag, R. V. & Aguinaldo, Z. -Z. A. 2017. Genetic differentiation and distribution of *Pyropia acanthophora* (Bangiales, Rhodophyta) in the Philippines. *Eur. J. Phycol.* 52:104–115.
- Excoffier, L. & Lischer, H. E. L. 2010. Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Mol. Ecol. Resour.* 10:564–567.
- Faria, R., Johannesson, K. & Stankowski, S. 2021. Speciation in marine environments: diving under the surface. *J. Evol. Biol.* 34:4–15.
- Ferrer, M. S. R., Gomez, R. N., Nacido, C. B., Moises, M. T. & Dumilag, R. V. 2019. Genetic diversity of Philippine *Gracilaria salicornia* (Gracilariaceae, Rhodophyta) based on mitochondrial COI-5P sequences. *Biologia* 74:599–607.
- Fraser, C. I., Zuccarello, G. C., Spencer, H. G., Salvatore, L. C., Carcia, G. R. & Waters, J. M. 2013. Genetic affinities between trans-oceanic populations of non-buoyant macroalgae in the high latitudes of the Southern Hemisphere. *PLoS ONE* 8:e69138.
- Gabriel, D., Draisma, S. G. A., Sauvage, T., Schmidt, W. E., Schils, T., Lim, P. -E., Harris, D. J. & Fredericq, S. 2016. Multilocus phylogeny reveals *Gibsmithia hawaiiensis* (Dumontiaceae, Rhodophyta) to be a species complex from the Indo-Pacific, with the proposal of *G. eilatensis* sp. nov. *Phytotaxa* 227:1–20.
- Guo, W., Banerjee, A. K., Ng, W. L., Yuan, Y., Li, W. & Huang, Y. 2020. Chloroplast DNA phylogeography of the Holly mangrove *Acanthus ilicifolius* in the Indo-West Pacific. *Hydrobiologia* 847:3591–3608.
- Haditjar, Y., Putri, M. R., Ismail, N., Muchlisin, Z. A., Ikhwan, M. & Rizal, S. 2020. Numerical study of tides in the Malacca Strait with a 3-D model. *Heliyon* 6:e04828.
- Kalyaanamoorthy, S., Minh, B. Q., Wong, T. K. F., von Haeseler, A. & Jermin, L. S. 2017. ModelFinder: fast model selection for accurate phylogenetic estimates. *Nat. Methods* 14:587–589.
- Kantachumpoo, A., Uwai, S., Noiraksar, T. & Komatsu, T. 2014. Levels and distribution patterns of mitochondrial *cox3* gene variation in brown seaweed, *Sargassum polycystum* C. Agardh (Fucales, Phaeophyceae) from South-east Asia. *J. Appl. Phycol.* 26:1301–1308.
- Katoh, K., Misawa, K., Kuma, K. -I. & Miyata, T. 2002. MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Res.* 30:3059–3066.
- King, R. J. & Puttock, C. F. 1989. Morphology and taxonomy of *Bostrychia* and *Stictosiphonia* (Rhodomelaceae/Rhodophyta). *Aust. Syst. Bot.* 2:1–73.
- Lewmanomont, K., Wongrat, L. & Supanwanid, C. 1995. *Algae in Thailand*. Integrated Promotion Technology Co., Ltd., Bangkok, 334 pp.
- Ludt, W. B. & Rocha, L. A. 2015. Shifting seas: the impacts of Pleistocene sea-level fluctuations on the evolution of tropical marine taxa. *J. Biogeogr.* 42:25–38.
- Minh, B. Q., Schmidt, H. A., Chernomor, O., Schrempf, D., Woodhams, M. D., von Haeseler, A. & Lanfear, R. 2020. IQ-TREE 2: new models and efficient methods for phylogenetic inference in the genomic era. *Mol. Biol. Evol.* 37:1530–1534.
- Muangmai, N., Fraser, C. I. & Zuccarello, G. C. 2015a. Contrasting patterns of population structure and demographic history in cryptic species of *Bostrychia intricata* (Rhodomelaceae, Rhodophyta) from New Zealand. *J. Phycol.* 51:574–585.
- Muangmai, N., Preuss, M., West, J. A. & Zuccarello, G. C. 2022. Cryptic diversity and phylogeographic patterns of the *Bostrychia intricata* species complex (Rhodomelaceae, Rhodophyta) along the coast of southeastern Australia. *Phycologia* 61:27–36.
- Muangmai, N., Preuss, M. & Zuccarello, G. C. 2015b. Comparative physiological studies on the growth of cryptic species of *Bostrychia intricata* (Rhodomelaceae, Rhodophyta) in various salinity and temperature conditions. *Phycol. Res.* 63:300–306.
- Muangmai, N., von Ammon, U. & Zuccarello, G. C. 2016. Cryptic species in sympatry: nonrandom small-scale distribution patterns in *Bostrychia intricata* (Cerami-

- les, Rhodophyta). *Phycologia* 55:424–430.
- Muangmai, N., West, J. A. & Zuccarello, G. C. 2014. Evolution of four Southern Hemisphere *Bostrychia* (Rhodomelaceae, Rhodophyta) species: phylogeny, species delimitation and divergence times. *Phycologia* 53:593–601.
- Nguyen, V. X., Detcharoen, M., Tuntiprapas, P., Soe-Htun, U., Sidik, J. B., Harah, M. Z., Prathep, A. & Papenbrock, J. 2014. Genetic species identification and population structure of *Halophila* (Hydrocharitaceae) from the Western Pacific to the Eastern Indian Ocean. *BMC Evol. Biol.* 14:92.
- Ninwichian, P. & Klinbunga, S. 2020. Population genetics of sandfish (*Holothuria scabra*) in the Andaman Sea, Thailand inferred from 12S rDNA and microsatellite polymorphism. *Reg. Stud. Mar. Sci.* 35:101189.
- Palmer, D. R. 2004. Phylogeography and population genetic structure of the dugongs in Thailand. M.S. thesis, San Jose State University, San Jose, CA, 61 pp.
- Panithanarak, T. 2020. Phylogeography of three commercially important seahorses (genus *Hippocampus*) in Thai waters: an implication from collective sequence data. *J. Fish. Environ.* 44:1–15.
- Payo, D. A., Colo, J., Calumpong, H. & De Clerck, O. 2011. Variability of non-polar secondary metabolites in the red alga *Portieria*. *Mar. Drugs* 9:2438–2468.
- Payo, D. A., Leliaert, F., Verbruggen, H., D'hondt, S., Calumpong, H. P. & De Clerck, O. 2013. Extensive cryptic species diversity and fine-scale endemism in the marine red alga *Portieria* in the Philippines. *Proc. R. Soc. B Biol. Sci.* 280:20122660.
- Pongparadon, S., Zuccarello, G. C., Phang, S. -M., Kawai, H., Hanyuda, T. & Prathep, A. 2015. Diversity of *Halimeda* (Chlorophyta) from the Thai–Malay Peninsula. *Phycologia* 54:349–366.
- Poommouang, A., Kriangwanich, W., Buddhachat, K., Brown, J. L., Piboon, P., Chomdej, S., Kampuansai, J., Mekchay, S., Kaewmong, P., Kittiwattanawong, K. & Ng-anvongpanit, K. 2021. Genetic diversity in a unique population of dugong (*Dugong dugon*) along the sea coasts of Thailand. *Sci. Rep.* 11:11624.
- Rambaut, A. 2016. FigTree v1.4.3. Available from: <http://tree.bio.ed.ac.uk/software/figtree/>. Accessed Mar 27, 2021.
- Rizal, S., Damm, P., Wahid, M. A., Sundermann, J., Ilham-syah, Y., Iskandar, T. & Muhammad. 2012. General circulation in the Malacca Strait and Andaman Sea: a numerical model study. *Am. J. Environ. Sci.* 8:479–488.
- Rogers, A. R. & Harpending, H. 1992. Population growth makes waves in the distribution of pairwise genetic differences. *Mol. Biol. Evol.* 9:552–569.
- Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D. L., Darling, A., Höhna, S., Larget, B., Liu, L., Suchard, M. A. & Huelsenbeck, J. P. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst. Biol.* 61:539–542.
- Rozas, J., Ferrer-Mata, A., Sánchez-DelBarrio, J. C., Guirao-Rico, S., Librado, P., Ramos-Onsins, S. E. & Sánchez-Gracia, A. 2017. DnaSP 6: DNA sequence polymorphism analysis of large data sets. *Mol. Biol. Evol.* 34:3299–3302.
- Saengkaew, J., Muangmai, N. & Zuccarello, G. C. 2016. Cryptic diversity of the mangrove-associated alga *Bostrychia* (Rhodomelaceae, Rhodophyta) from Thailand. *Bot. Mar.* 59:363–371.
- Sathiamurthy, E. & Voris, H. K. 2006. Maps of Holocene Sea level transgression and submerged lakes on the Sunda Shelf. *Nat. Hist. J. Chulalongkorn Univ. Suppl.* 2:1–44.
- Seesamut, T., Jirapatrasilp, P., Sutcharit, C., Tongkerd, P. & Panha, S. 2019. Mitochondrial genetic population structure and variation of the littoral earthworm *Pontodrilus longissimus* Seesamut and Panha, 2018 along the coast of Thailand. *Eur. J. Soil Biol.* 93:103091.
- Suppapan, J., Sangthong, P., Songrak, A. & Supmee, V. 2021. Population genetic structure of hard clam (*Meretrix lyrata*) along the Southern coast of Thailand. *Biodiversitas* 22:2489–2496.
- Tajima, F. 1989. Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics* 123:585–595.
- Tanabe, A. S. 2011. Kakusan4 and Aminosan: two programs for comparing nonpartitioned, proportional and separate models for combined molecular phylogenetic analyses of multilocus sequence data. *Mol. Ecol. Resour.* 11:914–921.
- Teske, P. R., von der Heyden, S., McQuaid, C. D. & Barker, N. P. 2011. A review of marine phylogeography in southern Africa. *S. Afr. J. Sci.* 107:43–53.
- Vieira, C., Steen, F., D'hondt, S., Bafort, Q., Tyberghein, L., Fernandez-García, C., Wysor, B., Tronholm, A., Mattio, L., Payri, C., Kawai, H., Saunders, G., Leliaert, F., Verbruggen, H. & De Clerck, O. 2021. Global biogeography and diversification of a group of brown seaweeds (Phaeophyceae) driven by clade-specific evolutionary processes. *J. Biogeogr.* 48:703–715.
- Wee, A. K. S., Takayama, K., Asakawa, T., Thompson, B., Sungkaew, S., Tung, N. X., Nazre, M., Soe, K. K., Tan, H. T. W., Watano, Y., Baba, S., Kajita, T. & Webb, E. L. 2014. Oceanic currents, not land masses, maintain the genetic structure of the mangrove *Rhizophora mucronata* Lam. (Rhizophoraceae) in Southeast Asia. *J. Biogeogr.* 41:954–964.
- Wichachucherd, B., Prathep, A. & Zuccarello, G. C. 2014.

- Phylogeography of *Padina boryana* (Dictyotales, Phaeophyceae) around the Thai-Malay Peninsula. *Eur. J. Phycol.* 49:313–323.
- Wyrski, K. 1961. *Physical oceanography of the Southeast Asian waters*. Scripps Institution of Oceanography, The University of California, La Jolla, CA, 195 pp.
- Zanolla, M., Altamirano, M., Carmona, R., De la Rosa, J., Souza-Egipsy, V., Sherwood, A., Tsiamis, K., Barbosa, A. M., Muñoz, A. R. & Andreakis, N. 2018. Assessing global range expansion in a cryptic species complex: insights from the red seaweed genus *Asparagopsis* (Florideophyceae). *J. Phycol.* 54:12–24.
- Zuccarello, G. C. & Martin, P. 2016. Phylogeography of the *Lessonia variegata* species complex (Phaeophyceae, Laminariales) in New Zealand. *Algae* 31:91–103.
- Zuccarello, G. C., Muangmai, N., Preuss, M., Sanchez, L. B., Loiseaux de Goër, S. & West, J. A. 2015. The *Bostrychia tenella* species complex: morphospecies and genetic cryptic species with resurrection of *B. binderi*. *Phycologia* 54:261–270.
- Zuccarello, G. C., Sandercock, B. & West, J. A. 2002. Diversity within red algal species: variation in world-wide samples of *Spyridia filamentosa* (Ceramiaceae) and *Murrayella pericladus* (Rhodomelaceae) using DNA markers and breeding studies. *Eur. J. Phycol.* 37:403–417.
- Zuccarello, G. C. & West, J. A. 2003. Multiple cryptic species: molecular diversity and reproductive isolation in the *Bostrychia radicans*/*B. moritziana* complex (Rhodomelaceae, Rhodophyta) with focus on North American isolates. *J. Phycol.* 39:948–959.
- Zuccarello, G. C. & West, J. A. 2011. Insights into evolution and speciation in the red alga *Bostrychia*: 15 years of research. *Algae* 26:21–32.
- Zuccarello, G. C., West, J. A., Kamiya, M. & King, R. J. 1999. A rapid method to score plastid haplotypes in red seaweeds and its use in determining parental inheritance of plastids in the red alga *Bostrychia* (Ceramiaceae). *Hydrobiologia* 401:207–214.